

PONTIFÍCIA UNIVERSIDADE CATÓLICA DO PARANÁ ESCOLA DE CIÊNCIAS DA VIDA PROGRAMA DE PÓS-GRADUAÇÃO EM ODONTOLOGIA ÁREA DE CONCENTRAÇÃO CLÍNICA ODONTOLÓGICA INTEGRADA

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ASSOCIATION OF ORAL CONDITIONS AND POLYMORPHISMS IN THE *TNF* CLUSTER GENES WITH CHRONIC KIDNEY DISEASE AND MORTALITY

Curitiba 2020

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Tese apresentada ao Programa de Pós-Graduação em Odontologia da Pontifícia Universidade Católica do Paraná, como parte dos requisitos para obtenção do título de Doutor em Odontologia, Área de Concentração em Clínica Odontológica Integrada (Ênfase em Estomatologia).

Orientador: Prof.^a Dr.^a Paula Cristina Trevilatto

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TERMO DE APROVAÇÃO

VALÉRIA KRUCHELSKI HUK

ASSOCIAÇÃO DE CONDIÇÃO BUCAL E DE POLIMORFISMOS NOS GENES DO CLUSTER DO TNF COM DOENÇA RENAL CRÔNICA E MORTALIDADE

Tese apresentada ao Programa de Pós-Graduação em Odontologia da Pontificia Universidade Católica do Paraná, como parte dos requisitos parciais para a obtenção do Título de **Doutor em Odontologia** Área de Concentração em **Clínica Odontológica Integrada com Ênfase em Estomatologia**.

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1 ARTIGO 1 - VERSÃO EM INGLÊS

TITLE PAGE
Association of oral conditions and polymorphisms in the TNF cluster
genes with chronic kidney disease and mortality
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1 Abstract

Background: Chronic Kidney Disease (CKD) is a complex condition. Exacerbation
in the inflammatory mediators is associated with increased morbidity and mortality.
The aim was to investigate the association of oral condition and genetic
polymorphisms in the TNF cluster genes with risk to chronic kidney disease and the
impact of genetic variables on mortality.

7 Material and Methods: This is a case-control study consisted of 242 subjects 8 divided into 122 patients with CKD undergoing hemodialysis and 120 patients 9 without the disease. Sociodemographic and clinical parameters of oral health were 10 analyzed. Markers for full coverage in the TNF cluster genes were selected from 11 information on the International HapMap Project website and genotyped. The 12 hemodialysis patients' group was followed up for 5 years. For the multivariate 13 analysis, the binary logistic regression model was adjusted and variables with the 14 significance of p<0.20 were included. Survival analysis was done using the Kaplan-15 Meier model and compared using the Log-rank test. Values of p < 0.05 were 16 considered significant.

17 **Results:** Advanced age (*p*=0.000), male sex (*p*=0.000), smoking habit (*p*=0.002),

18 and the G allele of rs2229094 of the LTA gene (p=0.012) were associated with CKD.

19 LTA polymorphisms, rs2844482 in the additive and recessive models (p=0.016;

20 p=0.007 respectively) and rs2229094, in the recessive model (p=0.029), were

21 associated with mortality in hemodialysis patients.

Conclusions: An association between sociodemographic and genetic aspects with
 CKD was found. LTA gene polymorphisms had also an impact on mortality of CKD
 patients in hemodialysis.

25

Keywords: Genetic polymorphism; chronic kidney failure, oral health, lymphotoxin-alpha, tumor necrosis factor-alpha.

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- 29 30

1 1. Introduction

2 Chronic Kidney Disease (CKD) is a complex condition in which environmental and 3 genetic factors are involved¹⁻⁴. The chronic renal patient has physiological 4 alterations due to loss of renal function, leading to an increase in the production of 5 inflammatory cytokines, such as interleukin 1 and 6 (IL-1 and IL-6), tumor necrosis factor (TNF), C-reactive protein, and oxidative stress markers^{1,5}. However, the 6 7 systemic inflammatory burden in individuals with CKD may vary and cannot be 8 explained only by factors such as hemodialysis or clinical procedures^{3,6}. 9 Exacerbation in the production of inflammatory mediators has been associated with 10 increased morbidity and mortality in this group of patients⁴.

Several factors contribute to the presence of chronic inflammation in CKD patients, including poor oral health⁷ and genetic background². Poor oral health status is related to chronic kidney disease as a source of inflammation and contributing to complications that result in morbidity and mortality observed in endstage patients⁸.

In CKD, chronic inflammation is mediated by cytokines^{1,5}, low molecular 16 17 weight glycoproteins that participate in numerous immunological processes, 18 including pro and anti-inflammatory activities. The TNF cluster is composed of the 19 LTA, TNFA, and LTB genes, located on the short arm of 6 chromosome, which 20 encode the cytokines LT- α , TNF- α , and LT- β , respectively⁹. LT- α and LT- β are secreted by lymphocytes, the first acts locally as a mediator of the acute 21 22 inflammatory response, recruiting and activating neutrophils, but it is produced in small guantities ^{9,10}. The second is a transmembrane protein that acts as an anchor 23 24 for LT- α . Besides, LT- β may be an important regulator of the inflammatory 25 response⁹. TNF- α is secreted by macrophages and participates in numerous 26 processes of the immune-inflammatory response. When produced at high levels it 27 can cause excessive inflammation and tissue damage locally or systemically¹¹.

It has been suggested that polymorphisms may alter gene expression and/or
 cytokine production. Moreover, SNPs may influence the phenotype and outcome in
 CKD patients³. The investigation of genetic polymorphisms in the *TNF* cluster genes
 becomes interesting since those genes play an important role in the pro inflammatory and immunological response.

1 Single nucleotide polymorphisms in the LTA, TNFA, and LTB genes have 2 been associated with the development and progression of different diseases, such as cardiopathy associated with Chagas Disease¹², rheumatoid arthritis¹³, and 3 4 tuberculosis¹⁴. Further, SNPs in the TNFA gene were associated with CKD 5 complications ^{15,16} and death risk ¹⁷. This study hypothesized that oral condition and 6 polymorphisms in the TNF cluster genes could be related to risk and prognostic in 7 chronic kidney disease.

8 Thus, this study aimed to investigate the association of oral aspects and 9 polymorphisms in the TNF cluster genes with CKD and mortality in hemodialyzed 10 patients.

11

12 2. Material and Methods

13 2.1 Study population

14 The first part of this study comprised an observational, cross-sectional study with a 15 sample composed of 242 subjects from the Southern region of Brazil of both 16 genders. A sample was divided into case group, with 122 patients with stage 5 CKD 17 undergoing hemodialysis (HD), selected at the Pro-Renal Foundation of Curitiba 18 (Pro-renal) and control group, with 120 subjects without CKD [glomerular filtration] 19 rate > 90 mL/min/1.73m² estimated according to MDRD (*Modification of Diet in* 20 Renal Disease)], selected at the Dentistry Clinics of the Pontifical Catholic 21 University of Paraná (PUCPR). The data were collected in the period from August 22 2004 to April 2008 (around sixty months).

23 Subjects were not included when had chronic use of anti-inflammatory drugs, 24 HIV infection, immunosuppressive chemotherapy, systemic active infection, current 25 or lactating pregnancies, with diseases of hard or soft oral tissues (except caries 26 and periodontal disease), with use of orthodontic appliances, and with the presence 27 of gingivitis and/or necrotizing ulcerative periodontitis.

28

Informed consent was obtained from all individual participants included in the 29 study, with the protocol approved by the Research Ethics Committee of PUCPR 30 (CAAE 25141813.4.0000.0020).

31

32 2.2 Clinical and sociodemographic parameters All patients answered a questionnaire to inform the baseline parameters such as
 age, sex, smoking habit, the main cause of CKD, and systemic conditions.

3

4 2.3 Oral condition parameters

The following dental health parameters were evaluated: dentist visit frequency,
flossing, toothbrushing frequency, plaque index (PI)¹⁸, calculus index (CI)¹⁹, gingival
index (GI)¹⁸, presence of periodontal disease (PD), mobility and xerostomia.

8 Dentist visit frequency was registered as quarterly/semester or 9 annually/rarely. Flossing was answered by the patient as "yes" or "no". 10 Toothbrushing was answered by the patients as less than three times a day or 11 equal/more than three times a day. For PI, CI, and GI, each tooth received a score 12 ranging from 0 to 3. The scores were recorded, added, and divided by the number 13 of teeth to obtain an individual index for each patient. The average for the indices 14 was carried out within the groups and later dichotomized for better visualization of 15 the results. The diagnosis of chronic periodontal disease (PD) was made based on clinical parameters²⁰. Mobility was verified by the evaluator and recorded as "yes" 16 17 or "no".

18

19 2.4 DNA collection and purification

20 DNA was obtained from buccal epithelial cells²¹. The participants did a mouthwash 21 with 5 mL of 3% glucose solution for 1 minute. Then, a sterile wood spatula was 22 used to scrape the buccal mucosa. Oral epithelial cells were pelleted by 23 centrifugation at 2000 rpm for 10 minutes. The supernatant was discarded, and the 24 cell pellet resuspended in 1.3 ml of extraction buffer [10 mM Tris-HCI (pH 7.8), 5 25 mM ethylenediaminetetraacetic acid (EDTA), and 0.5% sodium dodecyl sulfate 26 (SDS)]. Ten microliters of proteinase K (20 mg/ml) were added to the solution, and 27 this was incubated overnight at 65°C. DNA was purified with 10 M ammonium 28 acetate and 1 mM EDTA²².

29

30 2.5 Selection of markers and genotyping

Tag SNP markers, which capture the whole gene information, were selected according to the information available on the *International HapMap Project* website.

33 The release 24/phase 2_Nov08 was used, and the selected markers showed a

minimal allelic frequency of 5% in the African population (YRI). A cutoff of r²≥0.8 of
linkage disequilibrium (LD) was used. After applying the above criteria, six tag SNPs
for *LTA* (rs2844482, rs2009658, rs2071590, rs2229094, rs2516312, rs3093542),
two for *TNFA* (rs1800629 and rs2228088), and one for *LTB* (rs3093553) were
selected.

6 The tag SNPs were genotyped using polarized fluorescence (ABI 7500
7 platform) by real-time PCR technique using the TaqMan[™] Genotyping Master Mix
8 (Applied Biosystems 7500 Real-Time PCR System).

9

10 2.6 Survival analysis

11 In the second part of this study, the case group patients (n=117) were followed 12 prospectively for 5 years. During this prospective follow-up data were collected only 13 for the primary endpoint (mortality due to any causes) or end of the study. Five 14 patients had an incomplete dataset and, therefore, they were excluded from the 15 analysis.

16

17 2.7 Statistical analysis

18 The Haploview 4.2 software was used to estimate the Hardy-Weinberg equilibrium,

to establish the associated allele or reference allele for each tag SNP, and to
analyze the LD between the SNPs of the *LTA*, *TNFA*, and *LTB* genes.

SPSS version 20.0 was used for statistical analysis. The nominal variables were expressed in number and frequency and analyzed by Pearson's chi-square test. Continuous variables were expressed as mean and standard deviation using the t-test for independent samples.

Multivariate analysis was performed using the binary logistic regression model by the *backward conditional* method and variables with the significance of p<0.20 were included, the model was adjusted for variables with clinical impact. Univariate survival analysis was done using the Kaplan-Meier model. Survival in different genotype groups was compared using the Log-rank test. Values of p<0.05were considered significant.

31

32 2.8 Power Calculation

The calculation of the sample power was performed using the Genetic Power Calculation²³ setting as parameters: prevalence of hemodialysis patients of 0.061% in the Brazilian population²⁴, effect size of 2.0 for carrying one copy of the risk allele and 4.0 for carrying two copies of the risk allele, D'= 1, frequency of the rare allele of 30%, 1: 1 case and control ratio and level of significance of 0.05, with the power variation being dependent on the relative risk of the rare allele.

7 3. Results

8 3.1 Clinical and sociodemographic parameters

9 The clinical and sociodemographic characteristics of the sample were described in 10 Table 1. In this study, the mean age for the case group was 49.74±12.83 and for 11 the control group, 39.93 ± 9.51 . The variables higher age (p=0.000), male sex 12 (p=0.000), and smoking (p=0.000) were associated with CKD in the study population. The ethnic variable showed no significant difference between groups. 13 14 The main cause of CKD was chronic glomerulonephritis (33.6%), followed by 15 diabetic nephropathy (18.0%), and hypertensive nephropathy (11.5%). About 16 systemic conditions, most patients with CKD were hypertensive (73.0%) and had 17 hepatitis (23.3%), CVD (22.1%), and diabetes (21.3%).

18

19 3.2 Oral condition parameters

In the case group, 69 (56.6%) did not use dental floss (p=0.005) and 109 (89.3%) performed brushing less than three times a day (p=0.001). However, about dentist visit frequency, 66 renal patients (54.1%) reported visiting the dentist annually/rarely against 102 (85%) in the control group (p=0.000). The variables PI, CI, GI, and presence of PD did not show significant differences between the groups. In the CKD group, dental mobility was observed in 48 patients (39.3%) (p=0.003) (Table 2).

27

28 3.3 Genetic analysis

The genotypic frequencies of all the markers tested were in Hardy-Weinberg equilibrium in the control group. The distribution of the genotypic frequencies in the additive, dominant, and recessive models for the tag SNPs in the *LTA*, *TNFA*, and *LTB* genes is shown in table 3. Some failures in genotyping were observed and the individuals excluded from the analysis, justifying the variable sample number
 differently observed in each one of the genotypic models studied.

The LD map showed that rs2844482 and rs2009658 of the *LTA* gene had $r^2=1.00$, (Fig. 1). For the Brazilian population, only one of them should be genotyped in future analyzes because they are in perfect LD.

6

7 3.4 Multivariate analysis

8 The multivariate analysis by the backward conditional method was performed. The 9 variables: age (p=0.000), sex (p=0.000), smoking habits (p=0.002), and the 10 recessive model for the G allele of rs2229094 (p=0.012) of the *LTA* gene remained 11 associated with CKD (Table 4).

12

13 3.5 Survival analysis

The survival analysis showed there were 33 (28.2%) death events in the hemodialysis patients during de follow-up period. Overall, nine tag SNPs in the *TNF cluster* were analyzed and after the Log-rank test, only two polymorphisms (rs284482 and rs2229094) in the *LTA* gene were associated with mortality.

Figure 2 illustrates the only significant survival curves for each tag SNPs in the *LTA* gene. The G allele for the rs2229094 (in the recessive model) was associated (p=0.029) with mortality in hemodialysis patients during the follow-up. Furthermore, the rs2844482, allele T in the additive and recessive models respectively associated with mortality of CKD patients (p=0.016; p=0.007).

There was no significant association between *LTB* and *TNFA* polymorphisms
and mortality in this group of patients.

25

26 3.6 Results of power calculation

The analysis of the genetic power of the sample is directly dependent on the frequency of the risk allele. Thus, for the risk allele frequency of 0.3, the sample power was 95% (detailed in Table 5).

1 4. Discussion

Chronic kidney disease is a complex condition in which different factors interact for
development and progression. Inflammation is a known and well-established risk
factor, directly associated with the prognosis of patients^{3,4}.

5 In this study, CKD patients were approximately ten years older (49.7) than 6 the control group (39.9) and were mostly male. Although the prevalence of CKD is 7 higher in females, men appear to progress faster to end-stage chronic kidney 8 disease²⁵. The influence of sex on the progression of CKD is not well understood 9 yet. However, some differences may be associated with the individual's biological 10 component²⁶.

11 This study reinforces the interaction between smoking and CKD. Smoking 12 accelerates the progression of renal disease and increases the risk for CVD²⁷. 13 Evidence shows up to 50% increased the risk for tobacco related CKD²⁸. The habit 14 results in renal fibrosis, consequently contributing to a rapid decline in renal 15 function²⁹. These changes due to smoking increase the risk of mortality in patients 16 with CKD, due to cardiovascular and other causes²⁷.

17 In the univariate analysis, the results showed that there were low rates of 18 tooth brushing and flossing in the studied population. The status of oral health in 19 patients with CKD has already been reported³⁰. However, the increased frequency 20 of visits to the dentist observed, different from what the literature shows, is justified 21 because they receive close dental care assistance at the Pro-Renal Foundation in 22 Curitiba/PR, where CKD patients perform their treatment. Frequent dental visits and 23 treatment of oral infectious loci have been reported as fundamental to reduce 24 systemic inflammatory conditions in this particular group of patients^{7,31}. Further, the 25 results showed that 39.9% of patients with CKD presented mobility, probably 26 because of PD. The oral inflammatory disease, such as gingivitis and PD, are the 27 most common manifestations of poor oral health and are considered a focus of 28 inflammation^{8,32}. The impact of oral health on systemic health has been 29 discussed³³. Frequent dental visits and treatment of oral infectious loci to reduce 30 systemic inflammatory conditions in this group of patients have been reported as fundamental^{7,31}. Despite the lack of association of oral health variables in the 31 32 multivariate analysis, the results may reinforce the importance that understanding the interaction between oral health and systemic health is essential to maintain the
 quality of life of the patient on hemodialysis.

3 The pathophysiology of chronic kidney disease is well known and studied. 4 Cytokines link to their receptors located in the cell membrane and regulate the 5 transcription of several genes leading to changes in the renal tissues¹. LTA gene 6 encodes LT-α, a proinflammatory cytokine that shares a signaling network with 7 TNF- α^9 . The role of LT- α in the pathophysiology of kidney disease is little discussed 8 in the literature. In this study, the association between the allele G (in the recessive 9 model) of rs2229094 in the LTA gene and risk for CKD was found. This same tag SNP was also associated with different conditions^{34–37}, however, there is no other 10 11 study in the literature investigating the association of this polymorphism with CKD 12 in other populations. Further, there was no association of TNF and LTB genes 13 polymorphisms with CKD in the studied population.

It has been suggested that elevated levels in inflammatory markers predict
poor prognosis and mortality in dialysis patients³. Thus, polymorphisms in the *LTA*, *TNFA*, and *LTB* genes could be associated with mortality in this group of patients.
After the survival analysis, the polymorphisms rs2844482 and rs2229094, in the *LTA* gene, are described for the first time as risk factors for mortality in hemodialysis
patients.

However, this study has limitations. The frequency of the rarer alleles was low for some tag SNPs. Although the calculation of genetic power shows that the sample size is adequate, the replication of this study in other populations is encouraged to better elucidate the role of these genes influencing the susceptibility to CKD and the outcome in patients on hemodialysis.

It was concluded that sociodemographic aspects, smoking habit, and rs2229094 (allele G, in the recessive model) of the *LTA* gene were associated with CKD in the studied Brazilian population. Also, the polymorphisms rs2844482 and rs2229094 of the *LTA* gene were associated with mortality in hemodialysis patients and might be suggested as markers of bad prognosis for CKD.

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1 **Declarations**

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6

7 Conflict of interest

8 The authors declare that they have no conflict of interest.

9

10 Availability of data and material

- 11 Data will be made available upon request
- 12

13 Ethics approval

14 Informed consent was obtained from all participants included in the study, with the protocol approved 15 by the Research Ethics Committee of PUCPR (CAAE 25141813.4.0000.0020). The study was 16 performed by the ethical standards as laid down in the 1964 Declaration of Helsinki and its later 17 amendments or comparable ethical standards.

18

19 Consent

20 Informed consent was obtained from all individual participants included in the study.

21

22 Authors' contributions

23 All authors contributed to the study conception and design. The first author, Valéria Kruchelski Huk 24 de Andrade drafted the article and participated in the interpretation of data. The second author 25 Márcia Olandoski analyzed data and, also, participated in the interpretation of data. The authors 26 Miguel Carlos Riella and Roberto Pecoits-Filho provided intellectual content of critical importance to 27 the work described, especially about renal disease and diagnosis. Cleber Machado de Souza 28 participated in the conception and design of the study and revised the article. The last author, Paula 29 Cristina Trevilatto, provided intellectual content of critical importance to the work and revised the 30 final version. All authors read and approved the final manuscript and are accountable for all aspects 31 of this study.

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Tables

 Table 1 Clinical and sociodemographic variables for groups case and control.

General Data	Case (n=122)	Control (n=120)	p-value	OR (CI 95%)
Age ^a	49.74 ± 12.83	39.93 ± 9.51	0.000*	-
Sex ^b				
Male	81 (66.4)	38 (31.7)	0.000**	2.04 (1.54-2.69)
Female	41 (33.6)	82 (68.3)		
Ethnic Group ^b				
Cauca	si 87 (71.3)	94 (78.3)	0.208**	1.45 (0.81-2.61)
Non-caucasian	35 (28.7)	26 (21.7)		
Smoking habits ^b				
Yes	28 (23.0)	8 (6.7)	0.000**	4.17 (1.81-9.58)
No	94 (77.0)	112 (93.3)		
Main cause of CKD ^b				
Chronic glomerulonephritis	41 (33.6)	0 (0.0)	0.000**	-
Diabetic nephropathy	22 (18.0)	0 (0.0)	0.000**	-
Hypertensive Nephropathy	14 (11.5)	0 (0.0)	0.000**	-
Others	40 (16.4)	0 (0.0)	0.000**	-
Systemic condition ^b				
Anemia	41 (33.6)	3 (2.5)	0.000**	
Hypertension	87 (71.3)	2 (1.7)	0.000**	-
Hepatitis	28 (23.0)	2 (1.7)	0.000**	-
Cardiovascular Disease	27 (22.1)	2 (1.7)	0.000**	-
Diabetes	26 (21.3)	2 (1.7)	0.000**	-
^a Mean ± Standard Deviation				

^bAbsolute Number (%)

OR: Odds Ratio

CI (confidence interval) *p-value for T test for independent samples **p-value for Pearson *qui*-square test

1	Table 2 (Dral	condition	variables	for	groups	case	and	control	I.
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General Data	Case (n=122)	Control (n=120)	p-value	OR (CI 95%)
Dentist Visit Frequency ^b				
Quarterly/Semester	56 (45.9)	18 (15.0)	0.000*	0.20 (0.11-0.39)
Annualy/rarely	66 (54.1)	102 (85.0)		
Flossing ^b				
Yes	53 (43.4)	74 (61.7)	0.005*	2.09 (1.25-3.50)
No	69 (56.6)	46 (38.3)		
Toothbrushing ≤ 3 times/day ^b	, , , , , , , , , , , , , , , , , , ,			
Yes	109 (89.3)	88 (73.3)	0.001*	3.04 (1.50-6.15)
No	13 (10.7)	32 (26.7)		
Plaque Index (PI) ^b	,	()		
< 1	34 (27 9)	33 (27 5)	0 949*	0 98 (0 55-1 72)
> 1	88 (72 1)	87 (72 5)	0.010	0.00 (0.00 1.12)
Calculus Index (CI) ^b	00 (72.1)	07 (72.0)		
	06 (79 7)	102 (95.0)	0.202*	1 52 (0 70 2 07)
=0	90(70.7)	102 (85.0)	0.203	1.55 (0.79- 2.97)
Sincipal Index (Cl)b	20 (21.3)	18 (15.0)		
Gingival index (GI) ⁵	00 (07 0)		0.440*	
< 1	82 (67.2)	75 (62.5)	0.443	0.81 (0.47-1.37)
≥ 1	40 (32.8)	45 (37.5)		
Presence of PD ^b				
Yes	73 (59.8)	58 (48.3)	0.073*	1.59 (0.95-2.65)
No	49 (40.2)	62 (51.7)		
Mobility ^b				
Yes	48 (39.3)	26 (21.7)	0.003ª	2.34 (1.33-4.13)
No	74 (60.7)	94 (78.3)		
Xerostomia ^b				
Yes	55 (45.1)	16 (13.3)	0.000*	2.69 (1.72-4.23)
No	67 (54.9)	104 (86.7)		
DMF-T Index ^a	34.00 ± 9.26	19.02 ± 6.93	0.121**	-
^a Mean + Standard Deviation				

Mean ± Standard Deviation

234567 8 ^b Absolute Number (%)

OR: Odds Ratio

CI (confidence interval) * p-value for Pearson qui-square test **p-value for T test for independent samples

9

10

11 **Table 3.** Genetic analysis of *LTA*, *LTB* and *TNFA* genes in the additive, dominant 12 and recessive models

Cono	dhCNDa	Constia Madal	Constynes	Group	os - n (%)	n voluo	OR (CL95%)
Gene	UDSNP*	Genetic Woder	Genotypes	Case	Control	p-value	
			CC	79 (64.8)	81 (68.1)		
		Additive	СТ	36 (29.5)	36 (30.3)	0.251*	-
			TT	7 (5.7)	2 (1.7)		
LTA	rs2844482	Dominant T	TT + CT	43 (35.2)	38 (31.9)	0 500*	1 16 (0 67 1 08)
		Dominant	CC	79 (64.8)	81 (68.1)	0.000	1.16 (0.67 -1.98)
		Decessive T	CC + CT	115 (94.3)	117 (98.3)	0.007*	0.00 (0.05 4.00)
		Recessive I	TT	7 (5.7)	2 (1.7)	0.097*	0.28 (0.05 -1.38)
			66	79 (66 4)	81 (68 1)		
		Additive	00 CG	34 (28.6)	36 (30 3)	0 353*	_
		Auditive		6 (5 0)	2(17)	0.555	-
1 7 1	re2009658			40 (33.6)	2 (1.7)		
LIA	132003030	Dominant C	60 + 00	70 (66 4)	91 (69.1)	0.782*	1.07 (0.62 -1.85)
				112 (05.0)	117 (09.2)		
		Recessive C	00+00	6 (5 0)	2(17)	0.150*	0.32 (0.06 -1.62)
				0 (5.0)	2 (1.7)		
			CC	61 (50.0)	48 (40.7)		
		Additive	CT	51 (41.8)	58 (49.2)	0.347*	-
	rs2071590 Deminent T		TT	10 (8.2)	12 (10.2)		
LTA		Dominant T	TT+CT	61 (50.0)	70 (59.3)	0 1/7*	0.68 (0.41 1.14)
		Dominant	CC	61 (50.0)	48 (40.7)	0.147	0.08 (0.41 -1.14)
		Pocossivo T	CC + CT	112 (91.8)	106 (89.8)	0 506*	1 26 (0 52 1 68)
		Recessive I	TT	10 (8.2)	12 (10.2)	0.596	1.20 (0.52 -1.00)
			AA	60 (49.2)	57 (47.9)		
		Additive	AG	46 (37.7)	55 (46.2)	0.113*	-
			GG	16 (13.1)	7 (5.9)		
LTA	rs2229094	Dominant G	GG + AG	62 (50.8)	62 (52.1)	0.842*	0.95 (0.57_1.57)
		Dominant	AA	60 (49.2)	57 (47.9)	0.042	0.33 (0.37-1.37)
		Recessive G	AA + AG	106 (86.9)	112 (94.1)	0.056*	1 / 3 (1 05 - 1 93)
		Recessive O	GG	16 (13.3)	7 (5.9)	0.000	1.40(1.00 - 1.00)
			AA	114 (94.2)	118 (99.2)		
		Additive	AG	7 (5.8)	1 (0.8)	0.066*	-
LTA	rs2516312		GG	-	-		
		Dominant G	GG + AG	7 (5.8)	1 (0.8)	0.066*	0.13 (0.01-1.14)
			AA	114 (94.2)	118 (99.2)	0.000	
	Recessive G	AA + AG	121 (100.0)	119 (100.0)	-	-	

			GG	-	-		
			GG	114 (93.4)	109 (93.2)		
		Additive	CG	7 (5.7)	8 (6.8)	0.584*	-
			CC	1 (0.8)	-		
LTA	rs3093542	Dominant G	GG + CG	121 (99.2)	117 (100.0)	0 326*	1 96 (0 73-2 22)
		Dominant O	CC	1 (0.8)	-	0.020	1.00 (0.70 2.22)
		Recessive G	CC + CG	8 (6.6)	8 (6.8)	0 931*	0 95 (0 34-2 63)
			GG	114 (93.4)	109 (93.2)	0.001	0.00 (0.04 2.00)
			AA	106 (87.6)	103 (86.6)		
		Additive	CA	15 (12.4)	16 (13.4)	0.809*	-
			CC	-	-		
LTB	rs3093553	Dominant C	CC + CA	15 (12.4)	16 (13.4)	0 809*	1 09 (0 51 – 2 33)
		Dominant	AA	106 (87.6)	103 (86.6)	0.000	1.00 (0.01 2.00)
		Recessive C	AA + CA	121 (100.0)	119 (100.0)	_	_
			CC	-	-		
			GG	122 (100.0)	118 (98.3)		
	rs2228088	Additive	GT	-	2 (1.7)	0.152*	-
			TT	-	-		
TNFA		Dominant T	TT + GT	-	2 (1.7)	0.152*	2 03 (1 78 – 2 31)
		Dominant	GG	122 (100.0)	118 (98.3)		2.00 (1.70 2.01)
		Recessive T	GG + GT	122 (100.0)	120 (100.0)	_	_
			TT	-	-		
			GG	100 (82.0)	98 (82.4)		
		Additive	AG	22 (18.0)	21 (17.6)	-	-
			AA	-	-		
TNFA	rs1800629	Dominant A	AA + AG	22 (18.0)	21 (17.9)	0.087*	0.97 (0.50 - 1.88)
		Dominant A	GG	100 (82.0)	98 (82.4)	0.307	0.97 (0.50 - 1.66)
		Beenerive A	GG + AG	119 (100.0)	122 (100.0)		
		Recessive A	AA	-	-	-	-

^a SNP identifier based on NCBI dbSNP
 ^a p-value for Pearson chi-square test
 OR Odds Ratio
 CI (confidence interval)
 The difference in n values refer to failure in genotyping

- 7 **Table 4.** Results after multivariate analysis using a binary logistic regression
- 8 model by the backward conditional method.
- 9

Variable	<i>p</i> -value*	OR (CI 95%)
Age	0.000	1.08 (1.05-1.11)
Sex	0.000	4.14 (2.22-7.72)
Smoking habit	0.002	4.66 (7.99-12.11)
Rs2229094RecG	0.012	3.86 (1.35-11.03)

10 *p-value after multivariate analysis using the binary logistic regression model

11 ÓR Odds Ratio

12 CI (confidence interval)

13

Table 5. The calculation of the sample power was performed using the Genetic Power Calculation (Purcell et al., 2003) setting as parameters: prevalence of hemodialysis patients of 0.061% in the Brazilian population (Brazilian Census of Nephrology, 2018), effect size from 2.0 for carrying one copy of the risk allele and 4.0 carrying two copies of the risk allele, D'= 1, frequency of the rare allele of 30%, 1:1 case and control ratio and level of significance of 0.05, with the power variation being dependent on the relative risk of the rare allele.

0.05 0.12 1526 0.1 0.31 429 0.2 0.76 132 0.3 0.95 70 *Power of the sample when the rarer allele frequency is 30% a n cases for 80% power when the rarer allele frequency is 30%		Allele Frequency	Power	N cases for 80% power
0.1 0.31 429 0.2 0.76 132 0.3 0.95 70 *Power of the sample when the rarer allele frequency is 30% *n cases for 80% power when the rarer allele frequency is 30%		0.05	0.12	1526
*Power of the sample when the rarer allele frequency is 30% * n cases for 80% power when the rarer allele frequency is 30%		0.1	0.31	429
*Power of the sample when the rarer allele frequency is 30% * n cases for 80% power when the rarer allele frequency is 30%		0.2	0.76	70
	*Power ª n case	0.3 of the sample when the rare s for 80% power when the ra	0.95 allele frequency is 30 arer allele frequency is	% 30%

40 Figures

Fig 1 Analysis of linkage disequilibrium (LD) between the selected *LTA*, *TNFA* and *LTB* tag SNPs in the studied population. The LD value was shown inside the
squares and the number indicates the ratio in percent (%). The intensity of
the color within the squares reflects the LD between two tag SNPs. A darker
intensity represents the largest LD among the tag SNPs (1.0), while light gray
and white regions represent lower LD (<1.0).





100 References

- Zoccali C, Vanholder R, Massy ZA, et al. The systemic nature of CKD. Nat
 Rev Nephrol. 2017;13(6):344–58.
- 103 2. Nasef NA, Mehta S, Ferguson LR. Susceptibility to chronic inflammation: an
 104 update. Arch Toxicol. 2017; 91(3):1131–41.
- Nordfors L, Lindholm B, Stenvinkel P. End-stage renal disease--not an equal
 opportunity disease: the role of genetic polymorphisms. J Intern Med.
 2005;258(1):1–12.
- Balakrishnan VS, Guo D, Rao M, et al. Cytokine gene polymorphisms in hemodialysis patients: Association with comorbidity, functionality, and serum albumin. Kidney Int. 2004;65(4):1449–60.
- 5. Oncel M, Akbulut S, Toka Ozer T, et al. Cytokines, adipocytokines and
 inflammatory markers in patients on continuous ambulatory peritoneal
 dialysis and hemodialysis. Ren Fail. 2016; 6049:1–5.
- 114 6. Rao M, Wong C, Kanetsky P, et al. Cytokine gene polymorphism and
 115 progression of renal and cardiovascular diseases. Kidney Int.
 116 2007;72(5):549–56.
- 117 7. Ruokonen H, Nylund K, Furuholm J, et al. Oral Health and Mortality in
 118 Patients With Chronic Kidney Disease. J Periodontol. 2017; 88(1):26–33.
- Akar H, Akar GC, Carrero JJ, Stenvinkel P, Lindholm B. Systemic
 consequences of poor oral health in chronic kidney disease patients. Clin J
 Am Soc Nephrol. 2011;6(1):218–26.
- Ware CF. Network communications: lymphotoxins, LIGHT, and TNF. Annu
 Rev Immunol. 2005;23:787–819.
- 124 10. Junt T, V. Ta, Harris N, et al. Expression of lymphotoxin beta governs
 immunity at two distinct levels. Eur J Immunol. 2006;36(8):2061–75.
- Tracey D, Klareskog L, Sasso EH, Salfeld JG, Tak PP. Tumor necrosis factor
 antagonist mechanisms of action: a comprehensive review. Pharmacol Ther.
 2008;117(2):244–79.
- 129 12. Pissetti CW, de Oliveira RF, Correia D, Nascentes GAN, Llaguno MM,
 130 Rodrigues V. Association Between the Lymphotoxin-Alpha Gene
 131 Polymorphism and Chagasic Cardiopathy. J Interf Cytokine Res.
 132 2013;33(3):130–5.

- 133 13. Zhang C, Zhao M, Liu J, et al. Association of lymphotoxin alpha polymorphism
 134 with systemic lupus erythematosus and rheumatoid arthritis : a meta-analysis.
 135 2015;398–407.
- 136 14. Correa PA, Gómez LM, Anaya JM. Polimorfismo del TNF-alpha en
 137 autoinmunidad y tuberculosis. Biomédica. 2004;24:43.
- 138 15. Singh K, Prasad KN, Mishra P, et al. Association of tumour necrosis factor-α
 139 polymorphism in patients with end stage renal disease. Nephrology.
 140 2015;20(6):387–91.
- 141 16. Prakash S, Sarangi AN, Tripathi G, Sharma RK, Agrawal S. Prediction of
 142 susceptible biomarkers for end stage renal disease among North Indians.
 143 Nephrology. 2016;21(7):592–600.
- 144 17. Jaber B. Cytokine gene promoter polymorphisms and mortality in acute renal
 145 failure. Cytokine. 2004 7;25(5):212–9.
- 146 18. Loe H. The Gingival Index, the Plaque Index and the Retention Index147 Systems. J Periodontol. 1967;38:610–6.
- 148 19. Greene J, Vermillion J. The Simplified Oral Hygiene Index. J am Dent Assoc.149 1964;68:7–13.
- 150 20. Lindhe, J., Ranney, R., Lamster, I., et al. Consensus Report: Chronic
 151 Periodontitis. Annals of Periodontology. 1999; 4(1), 38–38.
- 152 21. Trevilatto PC, Line SR. Use of buccal epithelial cells for PCR amplification of
 153 large DNA fragments. J Forensic Odontostomatol. 2000;18(1):6–9.
- Aidar M, Line SRP. A simple and cost-effective protocol for DNA isolation
 from buccal epithelial cells. Braz Dent J. 2007;18(2):148–52.
- Purcell S, Cherny SS, Sham PC. Genetic Power Calculator: design of linkage
 and association genetic mapping studies of complex traits. Bioinformatics.
 2003;19(1):149–50.
- 159 24. Sociedade Brasileira de Nefrologia. SBN 2017 Censo. Vol. 144, Sbn. 2018.
 160 p. 1–2.
- 161 25. Neugarten J, Acharya A, Silbiger SR. Effect of gender on the progression of
 162 nondiabetic renal disease: a meta-analysis. J Am Soc Nephrol.
 163 2000;11(2):319–29.
- 164 26. Carrero JJ, Hecking M, Chesnaye NC, Jager KJ. Sex and gender disparities165 in the epidemiology and outcomes of chronic kidney disease. Nat Rev

166 Nephrol. 2018;14(3):151–64.

- 167 27. Nakamura K, Nakagawa H, Murakami Y, et al. Smoking increases the risk of
 168 all-cause and cardiovascular mortality in patients with chronic kidney disease.
 169 Kidney Int. 2015;88(5):1144–52.
- 170 28. Xia J, Wang L, Ma Z, et al. Cigarette smoking and chronic kidney disease in
 171 the general population: a systematic review and meta-analysis of prospective
 172 cohort studies. Nephrol Dial Transplant. 2017;32(3):475–87.
- 173 29. Van Laecke S, Van Biesen W. Smoking and chronic kidney disease: seeing
 174 the signs through the smoke? Nephrol Dial Transplant. 2017;32(3):403–5.
- 175 30. Klassen JT, Krasko BM. The dental health status of dialysis patients. J Can
 176 Dent Assoc. 2002;68(1):34–8.
- 177 31. Proctor R, Kumar N, Stein A, Moles D, Porter S. Oral and Dental Aspects of
 178 Chronic Renal Failure. J Dent Res. 2005;84(3):199–208.
- 179 32. Craig R. Interactions between chronic renal disease and periodontal disease.
 180 Oral Dis. 2007;14(1):1–7.
- 181 33. Kane SF. The effects of oral health on systemic health. Gen Dent. 65(6):30–182 4.
- 183 34. Hardikar S, Johnson LG, Malkki M, et al. A population-based case-control
 184 study of genetic variation in cytokine genes associated with risk of cervical
 185 and vulvar cancers. Gynecol Oncol. 2016;23(10):1780–9.
- 186 35. Harmon QE, Engel SM, Wu MC, et al. Polymorphisms in Inflammatory Genes
 187 are Associated with Term Small for Gestational Age and Preeclampsia. Am
 188 J Reprod Immunol. 2014 May;71(5):472–84.
- Mahajan A, Tabassum R, Srechavali E, et al. Obesity-dependent association
 of TNF-LTA locus with type 2 diabetes in North Indians. J Mol Med.
 2010;88(5):515–22.
- 192 37. Esposito S, Bosis S, Orenti A, et al. Genetic polymorphisms and the
 193 development of invasive bacterial infections in children. Int J Immunopathol
 194 Pharmacol. 2016;29(1):99–104.
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199 Parecer do Comitê de Ética em Pesquisa

PUCPE	ASSOCIAÇÃO PARANAENSE DE CULTURA - PUCPR	Porma
	PARECER CONSUBSTANCIADO DO CEP	
DADOS D	D PROJETO DE PESQUISA	
l'Itulo da	198QUISA: ANÁLISE DA ASSOCIAÇÃO GENÉTICA E A SUSCETIBILIDADE À DOE E A DOENÇA PERIODONTAL	NÇA RENAL
Peequisa	Ior: CLEBER MACHDO DE SOLIZA	
frea Terr	Itica: Gerética Humana: (Trata-se de pesquisa envolvendo Genétika Humana que não nesessi élica (or parte da CONEP.);	a de análise
versio:		
CAAE: 2	141613.4.0000.0020	
Patroeina	for Principal: Associação Paranaense de Cutura - PUCPR	
DADOS	DPARECER	
vumero o	D Parecer: 554,900	
data da M	Hatoria: 12/03/2014	
Apresent	ção do Projeto:	
A emenda	indui os genes inteneucinas (1, 4, 6, 8, 10, 12 e 17), do tator de necrose Vene de inteneros deno para e la estes nossam ser avaitados duanto a sua associat	an com a
pencare	al cronica (DRC) e com a dienca periodontal (DP). la que os mesnos estão envolvo	osna
esposta I	nune e infamatória nestas duas doenças.	
Eda errei	da está vinculada ao projeto principal.	
Objetive	a Posquisa:	
Objetvo F	Imario:	
D objetive	do presente trabalho é investigar a associação entre polimorfismos nos genes das i	interleueinas
1, 4, 6, 8	 10, 12 e 17], do tator de necrose tumoral (TNF)e de Interferons (IFN), que estão el mine e informatida na deseas regal artículas e a deseas periodadal. 	nvolvidos na
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